Response of Rice (Oryza sativa) to the Application of Manganese in Makurdi, Benue State, Nigeria

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Abstract. A pot experiment was carried out at the Agronomy Teaching and Research Farm of the University of Agriculture Makurdi to determine the effect of manganese fertilizer on the growth and yield of rice. Treatment consists of five levels of Manganese (Mn) (0, 5, 10, 15 and 20 kg ha⁻¹) in form of MnSO₄. The treatments were laid out in a Completely Randomized Design (CRD) with three replications. The effect of Mn on the growth of rice indicate that there were no significant difference in the growth parameters measured with the exception of plant height at 8 weeks after planting and number of tillers at 4 weeks after planting. The tallest plants and highest number of tillers were obtained with 10 kg ha⁻¹ Mn. The effect of Mn on the yield of rice indicate that there was a significant difference in the yield parameters measured with the exception of number of grain per panicle and root weight at 4 weeks after planting. Manganese at the rate of 10 kg ha⁻¹ gave the highest grain yield (2,667 kg ha⁻¹) and highest dry matter weight. Levels of Mn above 10 kg ha⁻¹ led to yield decrease, therefore application of 10 kg ha⁻¹ Mn was recommended for optimum yield of rice.

Introduction

Micronutrients are chemical elements necessary for plant growth in only extremely small amount. Although required in minute quantities, they have the same agronomic importance as macronutrients and play vital roles in the growth of plants [1]. These metallic chemical elements include Zinc (Zn), Iron (Fe), Copper (Cu) and manganese (Mn), amongst others. Micronutrients improve the yield and the crop quality for cereals, corn, beans, forages, and oil seed [2]. The deficiency of micronutrients reduces performance and profitability in the plant [3].

Manganese is one of the main micronutrients, which plays important role in plant as a component of enzymes involved in photosynthesis and other processes. Manganese is part of an important antioxidant (superoxide dismutase) structure that protects plant cells by deactivating free radicals which can destroy plant tissue. It plays vital roles in photosynthesis, as a structural component of the Photosystem II water splitting protein. It also serves as electron storage and delivery to the chlorophyll reaction centers [4, 5].

Rice (Oryza sativa) is a staple food in many countries of Africa and other parts of the world. This is the most important staple food for about half of the human race [6]. Saka and Lawal [7] classified rice as the most important food depended upon by over 50 percent of the World population for about 80 percent of their food need. Due to the growing importance of the crop, FAO [8] estimated that annual rice production should be increased from 586 million metric tons in 2001 to meet the projected global demand of about 756 million metric tons by 2030.

The drive towards self-sufficiency in food production through the adoption of more scientific intensive agricultural systems has necessitated the evaluation of the nutrient status of soils and their role in crop production; most especially the micronutrients which had hitherto been neglected [9].

However, few investigations carried out so far have revealed micronutrient deficiency in some Nigerian savanna soils [10-12]. These deficiencies may have resulted from intensively cultivated soil with high nutrient-demanding crops, highly weathered rocks and leaching. Mustapha and Loks [13] reported that the use of new high yielding crop varieties which are nutrient demanding have unraveled micronutrient deficiencies in some Nigeria Savanna soils. Also the non-inclusion of these
important elements (Mn) in fertilization programs have led to its widespread deficiency hence, the need to assess the response of crops to the application of these nutrients, particularly Manganese which facilitates the production of carbohydrates and is required for optimum utilization of macro nutrients in plants.

The present research was undertaken to determine the growth and yield response of rice to the application of Manganese fertilizer.

**Materials and Methods**

The present investigation was in two parts; laboratory studies and pot experiment which were carried out in 2017. The laboratory studies include routine soil analysis and micronutrient determination and were carried out at the Advanced Analytical Soil Laboratory of the Department of Soil Science, University of Agriculture, Makurdi while the pot experiment was carried out at the Agronomy Teaching and Research Farm of the same institution which lies between Latitude 70° 47' N and Longitude 80° 36' E at an elevation of 82 m above sea level.

**Soil Sampling and Analysis**

Surface Soil samples were taken at a depth of 0–20 cm from Agronomy Teaching and Research Farm, Federal University of Agriculture Makurdi, taking into consideration all possible precautions prescribed for soil sampling [14]. The samples were brought to the laboratory, air dried and mixed together thoroughly and crushed to pass a 2 mm sieve.

**Sample Analysis**

The soil samples were analysed for the following parameters using standard procedures; pH was measured by glass electrode in a 1: 2 soil, water ratio. Exchange acidity was determined by the titration method [15]. Exchangeable bases were extracted with neutral ammonium acetate solution buffered at pH 7. Na and K in the extract were determined using flame photometer while Ca and Mg were determined by Atomic Absorption Spectrophotometer (AAS) [15]. Organic matter was determined by wet acid digestion [16], Total Nitrogen by the Kjeldahl digestion method, phosphorus by Bray-I procedure [17] particle size analysis by the hydrometer method of Bouyoucos [18]. The CEC was determined by neutral, 1N Ammonium acetate method while Base Saturation was calculated by dividing the sum of exchangeable bases by CEC and multiplying by 100. Mn was extracted using 0.1 N HCl and read with atomic absorption spectrophotometer. The DTPA extractant was used to extract available micronutrients and the values read on an Atomic Absorption Spectrophotometer [19].

**Pot Experiment**

A pot experiment was conducted at the Teaching and Research Farm of the University of Agriculture Makurdi, North Core using soil collected from the farm. Four (4) kg of the soil was weighed into perforated plastic pots of 5 litres capacity.

Treatment consists of five levels of Mn (0, 5, 10, 15 and 20 kg ha⁻¹) in the form of MnSO₄ which contains 27 % Mn. The treatments were laid out in a Completely Randomized Design (CRD) with three replications. Ten (10) seeds of rice (L 34 variety) obtained from Olam rice farm, Rukubi Nasarawa state, Nigeria were planted per pot and later thinned to 5 plants per pot two weeks after planting. Fertilizer equivalent to 80 kg N, 30 kg P₂O₅ and K₂O were applied to all the pots as basal application [20] and the Mn rates distributed accordingly to the pots at planting with the control treatment pots having no Mn application.

**Data Collection**

Data was collected on the following parameters:

i. Number of tillers at 4 and 6 weeks after planting
ii. Plant height at 4, 6, 8 and 10 weeks after planting
iii. Root weight at 4, 6 and 8 weeks after planting
iv. Leaf area at 4, 6, 8 and 10 weeks after planting  
v. Number of grains per panicle at harvest  
vi. Grain yield at harvest

**Data Analysis**

All the plant data collected were subjected to statistical analysis of variance (ANOVA) for the  
determination of significant treatment effects. Those that have significant effects were separated  
using Fisher's Least Significant Difference (FLSD).

**Results and Discussion**

**Physical and Chemical Properties of the Experimental Soil**

The physical and chemical properties of the experimental soils are presented in Table 1. The  
pH was slightly acidic (6.29), Textural class was clay loam with 56 g kg\(^{-1}\) sand, 36 g kg\(^{-1}\) silt  
and 74 g kg\(^{-1}\) clay. The soil was generally low in Phosphorus (3.54 mg kg\(^{-1}\)), Calcium (3.33 cmol kg\(^{-1}\)),  
Magnesium (1.36 cmol kg\(^{-1}\)), Potassium (0.12 cmol kg\(^{-1}\)) and Sodium (0.36 cmol kg\(^{-1}\)). The  
Nitrogen content was also very low (0.02 %). Organic matter was 3.08 %, the CEC was low with  
base saturation of 84 %. The micronutrient status of the soil include; Copper which was medium  
(0.72 mg kg\(^{-1}\)), iron was medium (3.5 mg kg\(^{-1}\)), Manganese was low (0.70 mg kg\(^{-1}\)) and the Zinc  
content was low (0.02 mg kg\(^{-1}\)).

The soil used for this research had low fertility status and was slightly acidic in nature. This  
could be due to the removal of basic cations from the surface of the soils to the lower depths [13,  
22-23]. Results show that organic carbon content was high [23] this is contrary to the low organic  
carbon values reported by Yaro *et al.* [24] for the Nigerian Savanna soils, and those reported by  
Mustapha and Nnalee [25] and Mustapha *et al.* [26] for soils in the Northern Guinea Savanna Zone  
of Nigeria.

<table>
<thead>
<tr>
<th>Parameters</th>
<th>Values</th>
</tr>
</thead>
<tbody>
<tr>
<td>pH(1:1)</td>
<td>6.29</td>
</tr>
<tr>
<td>Sand (g kg(^{-1}))</td>
<td>56</td>
</tr>
<tr>
<td>Silt (g kg(^{-1}))</td>
<td>36</td>
</tr>
<tr>
<td>Clay (g kg(^{-1}))</td>
<td>74</td>
</tr>
<tr>
<td>Textural class</td>
<td>Clay Loam</td>
</tr>
<tr>
<td>Organic C (%)</td>
<td>1.79</td>
</tr>
<tr>
<td>Organic matter (%)</td>
<td>3.08</td>
</tr>
<tr>
<td>N (%)</td>
<td>0.02</td>
</tr>
<tr>
<td>Available P (mg kg(^{-1}))</td>
<td>3.54</td>
</tr>
<tr>
<td>Ca (cmol kg(^{-1}))</td>
<td>3.33</td>
</tr>
<tr>
<td>Mg (cmol kg(^{-1}))</td>
<td>1.36</td>
</tr>
<tr>
<td>K (cmol kg(^{-1}))</td>
<td>0.12</td>
</tr>
<tr>
<td>Na (cmol kg(^{-1}))</td>
<td>0.36</td>
</tr>
<tr>
<td>CEC (cmol kg(^{-1}))</td>
<td>5.17</td>
</tr>
<tr>
<td>B.S (%)</td>
<td>84</td>
</tr>
<tr>
<td>Exch A</td>
<td>0.83</td>
</tr>
<tr>
<td>Cu(mg kg(^{-1}))</td>
<td>0.72</td>
</tr>
<tr>
<td>Fe(mg kg(^{-1}))</td>
<td>3.50</td>
</tr>
<tr>
<td>Mn(mg kg(^{-1}))</td>
<td>0.70</td>
</tr>
<tr>
<td>Zn(mg kg(^{-1}))</td>
<td>0.22</td>
</tr>
</tbody>
</table>
The high organic carbon content could be due to the fact that plant residues were usually left on the soil after harvest year after year without burning. The decomposition of these residues may have contributed to the high organic carbon content.

The Zn level in the soils was low (0.02 mg kg\(^{-1}\)) [23]. The values recorded for the soil falls below the 1.2–4.0 mg kg\(^{-1}\) reported by Kparmwang and Malgwi [27] for the soils in the Northern Guinea Savanna of Nigeria, 0.81 to 1.34 mg kg\(^{-1}\) reported for Ustults in Galambi District in Bauchi State, Nigeria [28] and 1.1 to 6.9 mg kg\(^{-1}\) reported for volcanic ash soils elsewhere in Tanzania [29]. It is pertinent to note that the Zn values obtained in this study fall below the critical 0.90 mg kg\(^{-1}\) given by Lombin [11]. The low Zn content may be due to the non-inclusion of micronutrients in fertilizer programs which may lead to its deficiency as reported by Nziguheba et al. [30].

The low value of Cu (0.72 mg kg\(^{-1}\)) obtained in this research is lower than the results obtained by Mustapha and Singh [28] for soils elsewhere in Galambi, Bauchi State, Nigeria in similar Agro-Ecology but are similar to the 0.81 to 0.26 mg kg\(^{-1}\) obtained by Mustapha et al. [31] in Gombe State. The low Cu content may be due to increased use of high analysis NPK fertilizers containing no or lower quantities of micronutrient.

The Fe in the soils was low (3.5 mg kg\(^{-1}\)). These values are much higher than the critical value (2.5 mg kg\(^{-1}\)) reported by Esu [23], but lower to the 12.40 - 45.1 mg kg\(^{-1}\) reported by Mustapha and Singh [28] for Ustults in similar agro ecology in Nigeria.

The available Mn in the soil is low (0.7 mg kg\(^{-1}\)). This implies that the soils contain Mn below the critical available range of 3 to 5 mg kg\(^{-1}\) reported by Lindsay and Norvell [19] and 1–5 mg kg\(^{-1}\) reported by Esu [23]. The values obtained for these soils is below those obtained for some Ustults in Bauchi, Nigeria (7.89–12.00 mg kg\(^{-1}\)) reported by Kparmwang [32] in similar Nigerian soils. The low content of available Mn in the soils may have resulted from intensive cultivation of the soil with new high nutrient-demanding crops as reported by Mustapha and Loks [13]. These crops remove micronutrients (Mn) from the soil without being replaced which may lead to its depletion.

**Effect of Manganese on the Growth of Rice**

The effect of Mn on the growth of rice (Table 2) indicate that there were no significant difference in the growth parameters measured with the exception of plant height at 8 weeks after planting and number of tillers at 4 weeks after planting.

At 8 weeks after planting, the tallest plants (5.63 cm) were obtained with 10 kg ha\(^{-1}\) Mn however this was not significantly different from plants obtained with 5 kg ha\(^{-1}\) Mn but was significantly taller than plants obtained with 0, 15 and 20 kg ha\(^{-1}\) Mn, which were not significantly different from themselves.

The highest number of tillers (3.67) was obtained with 10 kg ha\(^{-1}\) Mn at 4 weeks after planting which was significantly higher from the other treatments but there was no significant difference in number of tillers obtained with 0, 5, 15 and 20 kg ha\(^{-1}\) Mn.

No significant variation in growth parameters were noticed by the application of manganese (Mn) with the exception of plant height at 8 weeks after planting and number of tillers at 4 weeks after planting. This may be due to application manganese in the soil as reported by Qin [33] that rice responds to high Mn (about 0.5 mg/L) application with higher plant height, root length, and contents of chlorophyll a and chlorophyll b than at low Mn level. This is in contrast to reports by Prashad et al. [34] and Shanmugam and Veeraputran [35] that further increase in Mn rate was found to reduce the growth attributes with the minimum being observed in control.

The application of manganese significantly increased number of tillers only at 4 weeks after planting than without manganese this may be due to addition of Mn, this is similar to result obtained by Mousavi [36] who reported that rice fertilized with manganese at 4 weeks after planting had more productive tiller than without manganese this may be presumably caused by manganese which increased the content of chlorophyll in leaves of rice. Manganese played an important role in the production of chlorophyll and its presence was very important in photosynthesis [36].
Effect of Mn Fertilization on the Yield of Rice

The effect of Mn on the yield of rice (Table 3) indicated that there were no significant difference in the yield parameters measured with the exception of dry matter weight (DMW), grain yield and root weight (RW) at 8 weeks after planting.

The highest Dry matter weight (1.67 g) was obtained with 10 kg ha\(^{-1}\) Mn but there was no significant difference between 10 and 15 kg ha\(^{-1}\) Mn. However 10 kg ha\(^{-1}\) Mn was significantly different from what was obtained from the other treatments but no significant difference in the dry matter weight obtained with 0, 5 and 20 kg ha\(^{-1}\) Mn.

The highest grain yield (2,667 kg) per hectare was obtained with 10 kg ha\(^{-1}\) Mn. However, this was significantly different from what was obtained with the other treatments. There was no significant difference in the grain yield obtained with 0, 5, 15 and 20 kg ha\(^{-1}\) Mn.

Table 2. Effect of Mn Fertilizer on the Growth of Rice

<table>
<thead>
<tr>
<th>Treatment</th>
<th>Plant height (cm)</th>
<th>Number Tillers</th>
<th>Leaf Area (cm(^2))</th>
</tr>
</thead>
<tbody>
<tr>
<td>Mn</td>
<td>4WAP</td>
<td>6WAP</td>
<td>8WAP</td>
</tr>
<tr>
<td>0 kg ha(^{-1})</td>
<td>34.70</td>
<td>42.90</td>
<td>44.80</td>
</tr>
<tr>
<td>5 kg ha(^{-1})</td>
<td>34.50</td>
<td>46.70</td>
<td>48.00</td>
</tr>
<tr>
<td>10 kg ha(^{-1})</td>
<td>39.20</td>
<td>51.10</td>
<td>53.60</td>
</tr>
<tr>
<td>15 kg ha(^{-1})</td>
<td>32.30</td>
<td>42.20</td>
<td>44.20</td>
</tr>
<tr>
<td>20 kg ha(^{-1})</td>
<td>33.70</td>
<td>42.50</td>
<td>43.50</td>
</tr>
<tr>
<td>LSD(P&lt;0.05)</td>
<td>NS</td>
<td>NS</td>
<td>7.75</td>
</tr>
</tbody>
</table>

WAP = Weeks after planting, NS = Not significant

Table 3. Effect of Mn fertilizer on Yield of Rice

<table>
<thead>
<tr>
<th>Mn</th>
<th>DMW (g)</th>
<th>NGPP</th>
<th>RW4WAP (g)</th>
<th>RW8WAP (g)</th>
<th>Grain yield (kg ha(^{-1}))</th>
</tr>
</thead>
<tbody>
<tr>
<td>0 kg ha(^{-1})</td>
<td>0.87</td>
<td>46.7</td>
<td>0.02</td>
<td>0.09</td>
<td>1000</td>
</tr>
<tr>
<td>5 kg ha(^{-1})</td>
<td>0.67</td>
<td>56.7</td>
<td>0.02</td>
<td>0.12</td>
<td>1333</td>
</tr>
<tr>
<td>10 kg ha(^{-1})</td>
<td>1.67</td>
<td>112.3</td>
<td>0.04</td>
<td>0.08</td>
<td>2667</td>
</tr>
<tr>
<td>15 kg ha(^{-1})</td>
<td>1.33</td>
<td>95.7</td>
<td>0.05</td>
<td>0.16</td>
<td>1800</td>
</tr>
<tr>
<td>20 kg ha(^{-1})</td>
<td>1.00</td>
<td>70.0</td>
<td>0.05</td>
<td>0.16</td>
<td>1667</td>
</tr>
<tr>
<td>LSD(P&lt;0.05)</td>
<td>0.63</td>
<td>NS</td>
<td>NS</td>
<td>0.04</td>
<td>999.8</td>
</tr>
</tbody>
</table>

DMW = Dry Matter Weight, NGPP = Number of Grain per Panicle, RW = Root Weight

At 8 weeks after planting 15 and 20 kg ha\(^{-1}\) Mn gave the highest root weight with no significant difference between them while 0 and 10 kg ha\(^{-1}\) Mn have the least root weight which are not significantly different from each other. There was no significant difference in the root weight obtained with 0 and 5 kg ha\(^{-1}\) Mn at 8 weeks after planting.
There was a significant response in yield parameters of rice by the application of manganese (Mn) with the exception number of grain per panicle and root weight at 4 weeks of planting. This may be as a result of application of manganese soil content as reported by Dube et al. [37] from a field trial on rice that the plant biomass, panicle weight, grain weight, 1000-grain weight, chlorophyll a and b content and Hill reaction activity increased with increasing concentrations of Mn up to 0.55 mg L$^{-1}$ followed by a decrease with further increase in Mn.

The favorable effect on dry matter production might be due to higher leaf area per hill and leaf area index associated with high photosynthetic efficiency of rice plant leading to enhanced photosynthates accumulation and their translocation [38].

**Conclusion**

From the findings of this study, the experimental soil was low in most plant nutrients including micronutrients. The response of rice to manganese application is generally low in the growth parameters with the exception of plant height and number of tillers. However, significant differences were observed in the yield response of the crop to levels of Mn fertilization most especially in terms of root weight, grain yield and dry matter weight. For optimum yield of rice, 10 kg ha$^{-1}$ Mn should be included in rice production fertilization programmes.

**Conflict of Interest**

The authors declare that there is no conflict of interest.

**References**


