Trend of arsenic pollution and subsequent bioaccumulation in *Oryza sativa* and *Corchorus capsularis* in Bengal Delta

Sayan Bhattacharya^{1,3,*}, Gunjan Guha², Kaushik Gupta³,
Dhrubajyoti Chattopadhyay⁴, Aniruddha Mukhopadhyay⁵,
Uday Chand Ghosh³

¹Department of Environmental Studies, Rabindra Bharati University, Kolkata, India
*Phone: +919830950351

²School of Chemical and Bio Technology, Sastra University, Thanjavur, India

³Department of Chemistry, Presidency University, Kolkata, India

⁴B. C. Guha Centre for Genetic Engineering and Biotechnology, University of Calcutta, Kolkata, India

⁵Department of Environmental Science, University of Calcutta, Kolkata, India

*E-mail address: sayan_evs@yahoo.co.in

ABSTRACT

Oryza sativa Linn. (rice) and *Corchorus capsularis* Linn. (jute) are the two major crops of the Bengal basin. Both rice and jute are generally grown in submerged flooded conditions, where arsenic bioavailability is high in soil. The consumers of the edible parts from both plants therefore face an inevitable source of exposure to arsenic, with consequent accumulation and toxicity. The objective of the study was to observe the *in-vivo* temporal variation of arsenic bioaccumulation in the different parts of *O. sativa* and *C. capsularis*. Rice plant specimens (*Aman* rice, *Ratna* variety) of different age groups (1, 2 and 3 months old) were analyzed in HG-AAS for absorbed arsenic content in different parts. The accumulation of arsenic remained significantly high in the initial phase of growth, but decreased with time. Amount of arsenic bioaccumulation followed the decreasing order: root > basal stem > median stem > apical stem > leaves > grains in all the three age groups of the rice plant samples. *C. capsularis* followed a trend of arsenic bioaccumulation similar to *O. sativa*. *O. sativa* had more accumulation potential than *C. capsularis*, but *C. capsularis* showed much higher efficiency of arsenic translocation in the above ground parts. This is the first ever report of time-dependent decrease in arsenic bioaccumulation in *O. sativa* and *C. capsularis*. The contamination level can reach the grain part in significant amount and can cause health hazards in more severely arsenic affected areas. Intensive investigation on a complete food chain is urgently needed in the arsenic contaminated zones for further risk assessments.

Keywords: arsenic; rice; jute; bioaccumulation; Bengal Delta

1. INTRODUCTION

Arsenic (As) is a metalloid that poses serious environmental threats due to its behemoth toxicity and wide abundance. Arsenic contamination in groundwater has been reported in

Bangladesh, India, China, Taiwan, Vietnam, USA, Argentina, Chile, and Mexico (Bhattacharya *et al.* 2012). In many of these places the concentration has exceeded the permissible limit of 10 ppb (Bhattacharya *et al.* 2012). The Bengal basin is regarded to be the most acutely arsenic affected geological province in the world (Mukherjee *et al.* 2008). Bengal delta is one of the largest deltas of the world and accommodates an enormous volume of sediments deposited during the Tertiary and Quaternary periods (Bhattacharya *et al.* 2007). During the late Holocene period, marshy or swampy lowlands developed in several parts of the Bengal Delta Plain and formed the peat and other sediments rich in organic matter (Umitsu 1993).

Since groundwater arsenic contamination was first reported in year 1983 from 33 affected villages in four districts in West Bengal, the number of villages has increased to 3417 in 111 blocks in nine districts till 2008 in West Bengal alone (Bhattacharya *et al.* 2007). Chakraborti and his group analyzed over 1,25,000 water samples for studying arsenic contamination in the Bengal basin and reported that arsenic levels in the affected areas of the basin vary between <1 and 1300 mg/l (Chakraborti *et al.* 2002, 2004). The use of As contaminated groundwater for irrigation of crops in this region elevates arsenic concentration both in the surface soil and the plants (Meharg and Rahman 2003; Roychowdhury *et al.* 2005).

Oryza sativa Linn. (rice) and *Corchorus capsularis* Linn. (jute) are the two major crops of the Bengal basin. *O. sativa* is the paramount food crop in this part of the world. *C. capsularis*, on the other hand, is economically important for extraction of fibers and is also consumed as food by rural people (Shukla and Pai 2005). Previous studies have reported bioaccumulation of arsenic by *O. sativa* and *C. capsularis* in arsenic contaminated zones (Meharg and Rahman 2003; Das 2007). Both rice and jute are generally grown in submerged flooded condition, where arsenic bioavailability is high in soil (Duxbury and Panaullah 2007). Rice is much more efficient at assimilating arsenic into its grain than other staple cereal crops (Williams *et al.* 2007). Contamination of different varieties of rice by arsenic has been widely reported in North America, Europe, Taiwan, China, Bangladesh and India (Robberecht *et al.* 2002). In fact, food surveys on the daily arsenic intake in the US and Europe showed, second to fish products, rice is a major dietary source of arsenic (Robberecht *et al.* 2002; Schoof *et al.* 1999). The consumers of the edible parts from both plants therefore face an inevitable source of exposure to arsenic, with consequent accumulation and toxicity. The present study was aimed towards estimation of temporal dependence of arsenic bioaccumulation in the different parts of *O. sativa* and *C. capsularis* with reference to arsenic levels in the diverse strata of soil and in the groundwater used for irrigation.

2. MATERIALS AND METHODS

2. 1. Site of the study

The study was performed in and around Kalinarayanpur (23°22' N, 88°56' E), Nadia, West Bengal, India. The region, while being among the maximally affected zones of the country due to arsenic pollution (Chakraborti *et al.* 2002), is greatly involved in agriculture, particularly in the cultivation of *O. sativa* and *C. capsularis*. The agricultural system in this region is mostly dependent on irrigation with arsenic contaminated groundwater obtained from a depth ranging from 70 ft. to 600 ft. through tube-wells. Huge amount of groundwater is used for agricultural irrigation. Most of this groundwater is contaminated with arsenic, which is deposited in the soil in contact with the irrigation water throughout the year.

2. 2. Analysis of arsenic content in *O. sativa* and *C. capsularis*

Levels of arsenic in the two plants were determined by HG-AAS. The plants (grown in agricultural fields irrigated with arsenic contaminated water) were collected carefully without damaging the roots. Two age groups (1 month and 3 months old) were selected for studying the extent of arsenic bioaccumulation in rice and jute plants in the initial and final phase of growth. Following a thorough washing of all samples with water, roots of each one were measured to note their respective lengths.

Each plant specimen was analyzed for absorbed arsenic content in different parts: roots, stem (basal), stem (median), stem (apical), leaves and grains (only in *O. sativa*). These plant parts were separated carefully and dried in a hot air oven at 70 °C for 72 h. All the dried ground samples were stored in a desiccator until required for chemical analysis. Plant samples were digested by HNO₃–H₂O₂ (Tang and Miller 1991) to determine As concentrations. About 200 mg. of oven dried, ground plant material was weighed into a dry clean 75 ml digestion tubes. Five ml. of concentrated HNO₃ were then added to the tubes and allowed to stand overnight. The following day, the digesting tubes were placed on a heating block and the temperature raised to 0 °C. After adding three 1-ml aliquots of 30 % H₂O₂ to each tube the temperature was gradually raised to 120 °C and the samples were allowed to digest for 3 h, after which about 3-4 ml digest remained. The digests were cooled, diluted to 50 ml with deionized water and filtered through Whatman No. 42 filter paper into acid-washed plastic bottles. Arsenic concentrations in different parts of rice and jute plants were analyzed using a graphite furnace-atomic absorption spectrophotometer (GF-AAS), Perkin-Elmer. All glassware and plastic ware was previously acid washed in 2 % HNO₃, rinsed in deionised water and dried.

2. 3. Statistical analyses

All analyses were carried out in triplicates. Data were presented as mean ± standard deviation (SD). Statistical analyses were performed by one-way ANOVA to determine significant differences between groups at $P < 0.05$. Estimated correlations were tested for significance by Student's t-test at the same confidence limit. MATLAB ver. 7.0 (Natick, MA, USA), SPSS ver. 9.05 (Chicago, IL, USA) and Microsoft Excel 2007 (Roselle, IL, USA) were used for the statistical and graphical evaluations.

3. RESULTS AND DISCUSSIONS

The current study revealed that there was a time-dependent decline of arsenic content in the diverse parts of *O. sativa* (Fig. 1). In all the plants samples of two different age groups (i.e. 1 month and 3 months old plants), maximum arsenic accumulation was observed in the roots followed by stem parts, leaves and grains. The accumulation of arsenic remained significantly high in the initial phase (i.e. 1 month age); particularly in root zones it was 149.4 mg/kg, in stem basal part 22.5 mg/kg, in stem middle part 6.34 mg/kg, in stem apical part 2.78 mg/kg, in leaves 1.53 mg/kg and in the early phase of grains (apical parts in tillering stage) 0.29 mg/kg in average (Fig. 1). The possible reason behind it is the existence of oxidized zone around the rice root zone which helps to form oxidized iron plaque. Iron plaque can efficiently bind arsenic and can reduce its translocation to the above ground tissues (straw, husk and grain) of the plant (Norra *et al.* 2005). The microbial communities of

the rhizosphere can also solubilize ferric iron in the rhizosphere by exuding siderophores to the root-plaque interface, which is another possible reason for increased bioavailability of arsenic in rice plants (Kraemer 2004). The concentration of arsenic in rice roots can reach as high as 160 mg/kg. due to the formation of iron plaques, as reported in a previous study and with synchrony with the present work (Bhattacharya *et al.* 2007). It has also been reported that the accumulation of arsenic in rice plants is highest in the root zones and decrease significantly in the upper parts of the plant (Liu *et al.* 2004).

This was verified by the results of the present study, which showed a root > basal stem > median stem > apical stem > leaves > grains trend of arsenic accumulation in *O. sativa*. As the permissible limit of arsenic in rice plant grains is 1 mg/kg. according to WHO, it is quite alarming that such high concentration of arsenic has been found in rice. Interestingly, bioaccumulation of arsenic decreased drastically with time and in the mature 3 months old plants, the accumulation levels were much lower than that of the initial 1 month old phase. In the 3 months old rice roots, arsenic bioaccumulated 31.4 mg/kg, in stem basal part 7.72 mg/kg, in stem middle part 0.52 mg/kg, in stem apical part 0.43 mg/kg, in leaves 0.12 mg/kg and in the grains 0.07 mg/kg. in average respectively. In both growth phases, the grain parts accumulated arsenic well below the permissible limit. In a previous study, approximately 0.081 % and 1.57 % of arsenic are accumulated in rice grain from arsenic-deposited land and contaminated soil and about 0.029 and 0.133 mg of arsenic are accumulated in rice grain per kg. of plant and kg. of paddy, respectively in Nadia district (Roychowdhury 2008).

The arsenic concentrations in the edible parts of a plant depend on the availability of the soil arsenic and the accumulation and translocation ability of a plant (Huang *et al.* 2006). The study can indicate that the contamination level can reach the grain part in significant amount and can cause health hazards in more severely arsenic affected areas. Interestingly, Roychowdhury (2008) found that in Murshidabad, there was a substantial increase of arsenic accumulation in rice after cooking (in raw rice the range of arsenic was found to be 0.043-0.662 mg/kg whereas in the cooked rice the range was 0.198-1.03) which can indicate that arsenic can create further health hazards because of cooking rice in arsenic contaminated water.

Arsenic toxicity can reduce the rate of photosynthesis in rice plants, which, in turn can reduce the chlorophyll content, and can affect the growth and yield of rice (Rahman *et al.* 2007). Rice yield has been reported to decrease by 10 % when the concentration of arsenic in soil is as high as 25 mg/kg (Xiong *et al.* 1987). It has been shown that in T-Aman rice (*Oryza sativa* L.), the grain and straw yields were significantly reduced by the artificial introduction of arsenic in soil (Azad *et al.* 2009). Trivalent arsenic has adverse effects on the photosynthetic efficiency and growth of the rice seedlings because of the involvement of MAP kinase cascade along with ROS and NO production in arsenic induced stress signal transduction in plants (Rao *et al.* 2011). In our study, the extent of accumulation indicated the fact that it may affect the productivity and quality of rice plants in the arsenic-contaminated zones, which in turn can affect the economic growth of the Bengal delta, as rice is the main cash crop of this region.

Figure 1. Diverse levels of arsenic bioaccumulation in different parts of *O. sativa*. A distinct time-dependent decrease in the levels of accumulated arsenic has been observed. Data are given in mean $\pm$ SD for $n = 3$ samples ($P < 0.05$).

While a very few studies were done on the arsenic bioaccumulation in *O. sativa*, not much research has been performed on arsenic accumulation in *C. capsularis* till date. The current investigation found that *C. capsularis* followed a trend of arsenic bioaccumulation similar to *O. sativa*. Fig. 2 demonstrates that there was a significant temporal dependence of the decline of arsenic levels in the different parts of the plant.

In the initial phase of growth (i.e. in 1 month), in jute roots, arsenic bioaccumulated 8.701 mg/kg, in stem basal part 6.08 mg/kg, in stem middle part 5.71 mg/kg, in stem apical part 3.92 mg/kg, in leaves 2.07 mg/kg in average respectively. However, in the 3 month old plants, the accumulation level in all plant parts is with the permissible limit and showed no threat. The roots were found to accumulate the maximum amount of arsenic, similar to *O. sativa*, however, the level of such accumulation was about 15-25 times higher in *O. sativa*. Interestingly, when compared to rice plants, jute plants showed much higher efficiency of arsenic translocation in the above ground parts, as can be evident from Fig. 1 and 2 (the ratios of arsenic bioaccumulation in different parts of jute plants is much higher compared to rice plants).

Figure 2. Diverse levels of arsenic bioaccumulation in different parts of *C. capsularis*. A distinct time-dependent decrease in the levels of accumulated arsenic has been observed. Data are given in mean $\pm$ SD for $n = 3$ samples ($P < 0.05$).

Since all parts of both plants showed gradual decline in arsenic levels over three months, it was pertinent to hypothesize that there might be some physiological mechanisms working in those plants that may lower the arsenic level with increasing age.

An explanation of this phenomenon might be the presence of a potential detoxification mechanism involving complexation of arsenite and arsenate through production of thiol rich peptides phytochelatins (Hartley-Whitaker *et al.* 2001). There may also be some active processes which can help to reduce inorganic arsenic species to less toxic methylated arsenic compounds (Nissen and Benson 1982). Again, there might be an age-dependent increased efflux of arsenic species from the root into the rhizosphere (Sharples *et al.* 2000). It has been reported that in yeasts, arsenic tolerance is provided by three contiguous genes in the cluster ACR1, ACR2 and ACR3: ACR1 encodes a putative transcription factor; ACR2 encodes an arsenate reductase; and ACR3 encodes a plasma membrane AsIII-efflux transporter. This mechanism ensures As(V) reduction and its removal from the cytosol to the external medium (Tripathi *et al.* 2007). There is a possibility of the presence of such arsenic removal system in rice and jute plants which may be another possible reason in the decline of arsenic bioaccumulation with time.

4. CONCLUSIONS

The study underscored the fact that arsenic bioaccumulation is a function of availability of the metalloid as well as the physiological traits of the accumulating plants; the accumulation levels of arsenic in *O. sativa* being much higher in comparison to *C. capsularis*. Hence, intensive studies on the different crops grown in arsenic contaminated zones are the paramount priority. Additionally, the edible parts of the tuber crops grown in the arsenic contaminated zones are exposed continuously to soil and irrigation water contaminated with arsenic, which, in turn, can increase arsenic level in the tuber part. The extent of arsenic bioaccumulation in these edible plants should be monitored carefully.

Rice straw is often used as a cattle feed which represents another entry route of arsenic into the food chain, as rice straw typically contains much higher amount of arsenic than grains, as evident from our study. Cattle population also used to drink water contaminated with arsenic in those areas, which, in turn, can further increase the toxicity level in food chain (Abedin *et al.* 2002). Cattle manure is often used as fuel in household purposes, which can also increase the contamination risk (Pal *et al.* 2007). Besides, the dry straw often been used by people as fuel, which can release arsenic in air as oxides, and can cause pollution and health hazards. The overall scenario indicates clearly that the bioavailability of arsenic in rice and other edible plants must be addressed to understand the importance of arsenic exposure from these food sources. Intensive investigation on a complete food chain is urgently needed in the arsenic contaminated zones, which should be our priority in future researches.

References

- [1] Abedin MJ, Cresser MS, Meharg AA, Feldmann J., Cotter-Howells J (2002) Arsenic accumulation and metabolism in rice (*Oryza sativa* L.). *Environ Sci Technol* 36: 962-968.
- [2] APHA. 1985. Standard methods for the examination of water and wastewater. American Public Health Association, Washington, DC.
- [3] Azad MAK, Islam MN, Alam A, Mahmud H, Islam MA, Karim MR, Rahman M (2009) Arsenic uptake and phytotoxicity of T-aman rice (*Oryza sativa* L.) grown in the As-amended soil of Bangladesh. *Environmentalist* 29: 436–440.
- [4] Bhattacharya P, Mukherjee AB, Bundschuh J, Zevenhoven R., Loeppert R. (eds.) (2007) Arsenic in Soil and Groundwater Environment: Biogeochemical Interactions, Health Effects and Remediation. Elsevier Press, Amsterdam, UK.
- [5] Bhattacharya S, Gupta K, Debnath S, Ghosh UC, Chattopadhyay DJ, Mukhopadhyay A (2012) Arsenic bioaccumulation in rice and edible plants and subsequent transmission through food chain in Bengal basin: a review of the perspectives for environmental health. *Toxicol Environ Chem* 94: 429-441.
- [6] Chakraborti D, Rahman MM, Paul K, Chowdhury UK, Sengupta MK, Lodh D, Chanda CR, Saha KC, Mukherjee SC (2002) Arsenic calamity in the Indian subcontinent: what lessons have been learned? *Talanta* 58: 3-22.
- [7] Chakraborti D, Sengupta MK, Rahaman MM (2004) Groundwater arsenic contamination and its health effects in the Ganga–Meghna–Brahmaputra Plain. *J Environ Monitor* 6: 74-83.

-
- [8] Das DK (2007) Effects of arsenic-contaminated irrigation water, zinc and organic matter on the mobilization of arsenic in soils in relation to rice (*Oryza sativa* L.). In: Bhattacharya, P., Mukherjee, A. B., Bundschuh, J., Zevenhoven, R., Loeppert, R.H. (eds.). Arsenic in soil and groundwater environment: biogeochemical interactions, health effects and remediation. Elsevier, Maryland.
- [9] Duxbury JM, Panaullah G (2007) Remediation of arsenic for agriculture sustainability, food security and health in Bangladesh. Water Service, FAO, Rome.
- [10] Hartley-Whitaker J, Ainsworth G, Vooijs R, Bookum WT, Schat H, Meharg AA (2001) Phytochelatins are involved in differential arsenate tolerance in *Holcus lanatus*. *Plant Physiol* 126: 299-306.
- [11] Huang R, Gao S, Wang W, Staunton S, Wang G (2006) Soil arsenic availability and the transfer of soil arsenic to crops in suburban areas in Fujian Province, southeast China. *Sci Total Environ* 368: 531-541.
- [12] Kraemer S (2004) Iron oxide dissolution and solubility in the presence of siderophores. *Aquat Sci Res* 66: 3-18.
- [13] Liu WJ, Zhu YG, Smith SA, Smith SE (2004) Do iron plaque and genotypes affect arsenate uptake and translocation by rice seedlings (*Oryza sativa* L.) grown in solution culture? *J Exp Bot* 55: 1707-1713.
- [14] Meharg AA, Rahman MM (2003) Arsenic contamination in Bangladesh paddy field soils: implication for rice contribution to arsenic consumption. *Environ Sci Technol* 37: 229-234.
- [15] Mukherjee A, von Brömssen M, Scanlon BR, Bhattacharya P, Fryar AE, Hasan MA, Ahmed KM, Chatterjee D, Jacks G, Sracek O., *J Contam Hydrol* 99 (2008) 31-48.
- [16] Nissen P, Benson AA (1982) Arsenic metabolism in fresh tissue of arsenic by phytochelatins in plants. *Plant Physiol* 122: 793-801.
- [17] Norra S, Berner ZA, Agarwala P, Wagner F, Chandrasekharam D, Stüben D (2005) Impact of irrigation with As rich groundwater on soil and crops: a geochemical case study in West Bengal Delta Plain, India. *Appl Geochem* 20: 1890-1906.
- [18] Pal A, Nyack B, Das B, Hossain MA, Ahamed S, Chakraborti D (2007) Additional danger of arsenic exposure through inhalation from burning of cow dung cakes laced with arsenic as a fuel in arsenic affected villages in Ganga-Meghna-Brahmaputra plain. *J Environ Monitor* 9: 1067-1070.
- [19] Rahman MA, Hasegawa H, Rahman MM, Islam MN, Miah MAM, Tasmin A (2007) Effect of arsenic on photosynthesis, growth and yield of five widely cultivated rice (*Oryza sativa* L.) varieties in Bangladesh. *Chemosphere* 67: 1072-1079.
- [20] Rahman MA, Hasegawa H, Rahman MM, Rahman MA, Miah MA (2007) Accumulation of arsenic in tissues of rice plant (*Oryza sativa* L.) and its distribution in fractions of rice grain. *Chemosphere* 69: 942-948.
- [21] Rao KP, Vani G, Kumar K, Wankhede DP, Misra M, Gupta M, Sinha AK (2011) Arsenic stress activates MAP kinase in rice roots and leaves. *Arch Biochem Biophys* 506: 73-82.

-
- [22] Robberecht H, Van Cauwenbergh R, Bosscher D, Cornelis R, Deelstra H (2002) Daily dietary total arsenic intake in Belgium using duplicate portion sampling and elemental content of various foodstuffs. *Eur Food Res Technol* 214: 27-32.
- [23] Roychowdhury T, Tokunaga H, Uchino T, Ando M (2005) Effect of arsenic-contaminated irrigation water on agricultural land soil and plants in West Bengal, India. *Chemosphere* 55: 799-810.
- [24] Roychowdhury T (2008) Impact of sedimentary arsenic through irrigated groundwater on soil, plants, crops and human continuum from Bengal delta: special reference to raw and cooked rice. *Food Chem Toxicol* 46: 2856-2864.
- [25] Schoof RA, Yost LJ, Eickhoff J, Crecelius EA, Cragin DW, Meacher DM, Menzel DB (1999) A market basket survey of inorganic arsenic in food. *Food Chem Toxicol* 37: 839-846.
- [26] Sharples JM, Meharg AA, Chambers SM, Cairney JWG (2000) Mechanism of arsenate resistance in the Ericoid mycorrhizal fungus *Hymenoscyphus ericae*. *Plant Physiol* 124: 1327-1334.
- [27] Shukla SR, Pai RS (2005) Adsorption of Cu(II), Ni(II) and Zn(II) on modified jute fibres. *Bioresour Technol* 96: 1430-1438.
- [28] Tang T, Miller DM (1991) Growth and tissue composition of rice grown in soil treated with inorganic copper, nickel, and arsenic. *Commun. Soil Sci Plant Anal.* 22: 2037-2045.
- [29] Tripathi RD, Srivastava S, Mishra S, Singh N, Tuli R, Gupta DK, Maathuis FJM (2007) Arsenic hazards: strategies for tolerance and remediation by plants. *Trends Biotechnol* 25: 158-165.
- [30] Uemitsu M (1993) Late Quaternary sedimentary environments and landforms in the Ganges Delta. *Sediment Geol* 83: 177-186.
- [31] Williams PN, Vilada A, Deacon C, Raab A, Figuerola J, Green AJ (2007) Greatly enhanced arsenic shoot assimilation in rice leads to elevated grain levels compared to wheat and barley. *Environ Sci Technol* 41: 6854-6859.
- [32] Xiong XZ, Li PJ, Wang YS, Ten H, Wang LP, Song LP (1987) Environmental capacity of arsenic in soil and mathematical model. *Chinese J Environ Sci* 8: 8-14.