Synthesis, Characterization and Biological Evaluation of 3'-Benzoyl-5'-
(Furan-2-yl)-4'-Phenylspiro[indoline-3,2'-Pyrrolidin]-2-One Derivatives
and its Molecular Docking Studies

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Abstract. A series of new spirooxindole derivatives were synthesized via 1,3–dipolar
cycloaddition. All the synthesized compounds were evaluated for antimicrobial activity. In
antibacterial studies compound 4d demonstrated the most potent inhibitory activity (MIC =
12.5 lg/mL for K.pneumonia, B.cereus and S.typhi), which was compared with the positive control
streptomycin. In antifungal studies compound 4e demonstrated the most potent inhibitory activity
(MIC = 3.125 lg/mL for C.albicans and A.niger), which was related with the standard drug
ketaconazole. In addition, molecular modeling studies were also performed to disclose the binding
modes of the most active inhibitors to the amino acid residues that compose the active site of the
glucosamine-6-phosphate synthase and crystal structure of human lanosterol 14-alpha dimethylase
in complex with ketaconazole enzyme.

Introduction

Examining new bioactive promoters with the unimportant wide arrangement of built steps and
in significantly less time is a first rate dare to the researchers [1]. In any case, physicists have
opposed some other mission as far back as a long time in growing new techniques that are green,
specific and over the top yielding furthermore environmentally liberal. In doubtlessly
comprehended, the standard techniques incorporate the usage of multistep reaction groupings which
had been associated with low yields, absurd cost and inconvenience within the disconnection of the
stock. The use of multicomponent reactions (MCRs) has create as a present strategy which includes
three or extra straightforward and versatile particles united to rapidly introduce assistant
multifaceted nature and grouping, and offers sizable preferences over traditional direct sort union
[2]. Spirooxindoles are seen to have a wide arrangement of regular activities, which fuses
antimicrobial and antitumor, despite inhibitors of the human NKI receptor. Particularly,
spirooxindolepyrrolidine cross breed particles display neighborhood pain relieving results [3].
Despite this, spirotrypostatin has antimitotic houses with one of the little molecules, MDM2, in
preclinical change. As of late spirooxindole–pyrrolidine creamer particles have been exhibited to
have promising against TB relaxation movement [4, 5]. In light of the expansive natural activities of
these irritates, the mix of these iotas is a district of consistent studies. In classy, isatin and
auxiliaries are used as starting substances to guide 1,3-dipolar cycloadditions to yield spirooxindole
focus structures.[6] In perspective of the ease of rule, the imines created from isatin with an amino
acids or amines are reliably picked as huge 1,3-dipolar intermediates to react with specific
dipolarophiles, for example,β-nitrostyrenes, α,β-unsaturated esters, dienones, α,β - unsaturated
ketones, novel strong inhibitors of cutting edge glycation final item and electron-poor alkenes [7, 11].
A weighty part of the thought about 1,3-dipolar cycloadditions, β-nitrostyrenes are the most
broadly used dipolarophiles, in light of their application as suitable dipolarophiles for 1,3-dipolar
cycloaddition reactions to get prepared spirooxindoles. Chen and associates communicated their
revelations within the 1,3-dipolar cycloaddition reactions of isatin, an amino acids and b-
nitrostyrenes with emerge regioselectivity [12]. Perumal and partners said 1,3-dipolar cycloaddition
reactions of isatin, phenylglycine/proline/thiaproline with β-nitrostyrenes and surveyed those blends
for their in vitro activity towards Mycobacterium tuberculosis H37Rv (MTB) [13]. Finally, Peng-Fei Xu and partners suggested that an uneven 1,3-dipolar cycloaddition reaction of isatin, benzyl amine and β-nitrostyrenes with suitable selectivity in exceptional yield joins using steeply-esteemed chiral driving forces [14]. There's no fundamental system for the mix of spirooxindoles conveyed by the aggregate of benzyl amine, isatin and β-nitrostyrenes. Inferable from the natural significance of this tastefulness of blends, there is a need to add to a capable, fresh and speedy technique to overhaul the library of spirooxindoles. On this endeavor to get entry to the library of blends containing both spirooxindoles and pyrrolidine studs, we found the congruity of a microwave (MW) light approach to manage add to a gainful, supportive, over the top yield and quick fresh tradition.[15] In latest years, more highlight has been given to developing basic and fresh traditions for the mix of various trademark blends. Despite this, examination is in like way focused at the reaction in watery media as a result of precise homes like arranged openness, non-hurtful nature and wellbeing in overseeing.

**Experimental**

**General method**

All the reported melting points were taken in open capillaries and are uncorrected. IR spectra were recorded in AVATAR-330 FT-IR spectrophotometer (Thermo Nicolet) and only noteworthy absorption levels (reciprocal centimeters) are listed. The 1H and 13C NMR spectra at 400 and 100 MHz, respectively were obtained at room temperature using a Bruker 400 MHz NMR spectrometer (Bruker biospin, California, USA).

**General procedure for the synthesis of 3-benzoyl-5-(furan-2-yl)-4-phenylspiro[indoline-3,2-pyrrolidin]2-one derivatives (4a-g)**

An oven-dried hip flask was chilled under a stream of nitrogen and stimulating with a mixture of chalcones (1.0 mmol), isatin (1.0 mmol), furfuryl amine (1.0 mmol) in methanol (20ml). The mixture was refluxed until the vanishing of the preliminary materials as evidenced by the TLC (Scheme 2). After the reaction was over, the solvent was removed in vacuo and the residue was chromatographed on silica gel using hexane-ethyl acetate (7:3) as eluent to give the cycloadducts (4a-g) in tremendous yields.

**Molecular Docking Study**

Molecular docking experiment was carried out to study the exact binding location of ligand on protein. Molecular docking simulation was performed with the Argus Lab 4.0.1. The prepared 3D structures of 2VF5 and 3LD6 protein was downloaded from the protein data bank (see http://www.rcsb.org/pdb) and binding site was made by choosing “Making binding site for this protein” option. The ligand was then introduced and docking calculation was allowed to run using shape-based search algorithm and AScore scoring function. The scoring function is responsible for evaluating the energy between the ligand and protein target. Flexible docking was allowed by constructing grids over the binding sites of the protein and energy based rotation is set for that ligand group of atoms that do not have rotatable bonds. For each rotation, torsions and created and poses (conformation) are generated during the docking process. For each complex 10 independent runs were conducted and one pose was returned for each run. The best docking model was selected according to the lowest AScore calculated by arguslab and the most suitable binding conformation was selected on the basis of hydrogen bond interaction between the ligand and protein near the substrate binding site. The lowest energy poses indicate the highest binding affinity as high energy produces the unstable conformations. The resulting receptor model was saved to Brookhaven PDB file from the file the 2D and 3D interactions are viewed in discovery studio 4.1 versions.
Antimicrobial Assay

Collection of bacterial strains

The Clinical isolates of bacterial strains viz., K.pneumonia, B.cerus, S.typhi, E.coli and V. cholerae. The antifungal strains viz., Candida albicans, A.niger, A. Flavus, C. neoformans and F.oxysporum. These strains were obtained from the Department of Botany, Annamalai University, Annamalainagar, Tamil Nadu, India. The strains were inoculated on a sterile medium and sub-cultured on to Mueller Hinton Agar plates, these strains are maintained on agar slant at 4 °C.

Disc diffusion assay

Antibacterial and antifungal activity was performed by the disc diffusion method. About 1 mg/mL stock solution was prepared by dissolving the test compounds (7a-f) in 50% DMSO. The sterile paper disc with 6 mm diameter was impregnated with concentration of 200 mg/mL and the discs were placed in Mueller Hinton Broth for bacteria and Sabourauds dextrose broth for fungi. The plates were incubated at 37 °C for bacteria and 28 °C for fungi in incubator. The zone of inhibition for bacteria was visually examined at 37 °C for 24 h and fungi was visually examined at 28 °C for 72-96 h. Streptomycin was used as a standard positive control for bacteria and ketoconazole was used as a standard positive control for fungi under analogues conditions. All the tests were carried out in triplicate.

Minimum Inhibitory Concentration (MIC)

Dilution susceptibility testing method was used for MIC determination with reference to the cited literature. The test compounds were dissolved in 1 ml of chloroform. The different test concentrations of test compounds were 200 - 3.125 µg/ml. It was then serially diluted in to two folds. Wherein, 100 ml of sterile Mueller Hinton Broth for bacteria and Sabourauds Dextrose broth for fungus was decanted into each well of a sterile 96-well micro plate. Highest concentration of the test compounds added at 100 ml to the first well. After mixing of the above, 100 ml of the same was transferred to the second well and in this way; the dilution procedure was continued as a series of dilution of 200 - 3.125 µg/ml respectively. Inoculums solution at 5 ml was added to every well. Being incubated for 24 h at 37 ° C for bacteria and 28°C for fungi, the tubes were monitored for turbidity as growth and non-turbidity as no growth. The MIC values were interpreted as the highest dilution (lowest concentration) of the sample, which showed clear fluid with no development of turbidity. Bacterial and fungal growth was indicated by the measure of a white pellet on well bottom.

Results and Discussion

In the IR spectrum of 3'-benzoyl-5'-(furan-2-yl)-4'-phenylspiro[indoline-3,2'-pyrrolidin]-2-one, the -NH group of indolone moiety exhibited a sharp absorption peak at higher frequency at 3256 cm⁻¹ due to the presence of adjacent carbonyl group whereas the peak observed lower frequency at 3110 cm⁻¹ was assigned to -NH group of pyrrolidine ring. A strong absorption band at 1714 cm⁻¹ was due to indolone carbonyl stretching and a shoulder band at 1700 cm⁻¹ due to benzoyl carbonyl stretching. The aromatic and aliphatic C-H stretching absorption bands are observed in the region of 2853-3110 cm⁻¹. The observed imine and aromatic C-H stretching frequencies are the evidences for the formation of compounds 4a-g. ¹H NMR spectra of compounds 4a-g have been recorded in the solvent CDCl₃. The signals of the ¹H NMR spectra were assigned based on their positions, multiplicities and integral values. In ¹H NMR spectrum of the representative compound 4a, there was a multiplet in the region of 6.91-7.91 ppm corresponding to 14 protons are assigned to the aromatic protons. A broad doublet appeared at 4.55 ppm is assigned to H-5 proton due to its coupling with H-4 proton. A multiplet appeared at 5.54 ppm is assigned to H-3 and H-4 protons due to its coupling of H-3, H-4 and H-5 protons. A doublet appeared at 6.00 ppm is assigned to H-3″ due to its coupling with H-4″ proton. A broad singlet appeared at 6.13 ppm is assigned to H-4″ proton. A broad doublet appeared at 6.66 ppm is assigned to H-5″ proton due to its coupling with
H-4″ proton. From these observations we have confirmed the formation of synthesized compounds 4a-g. $^{13}$C-NMR spectrum exhibited the occurrence of two carbonyl carbons, one due to benzyol and another due to indolone moiety at 196.5 and 180.3 ppm respectively and the signals of the aromatic carbons are observed in the region of 142.5-108.2 ppm. The spiro carbon (C-2) of the pyrrolidine ring appeared as the most downfield signal at 72.2 ppm among all the carbons of this ring, while the peak at 57.0 ppm was regarded due to C-3 because of an adjacent carbonyl group of benzyol moiety. Further a signal observed at 54.5 ppm was assigned to C-4 carbon and another most upfield signal observed at 52.4 ppm was due to C-5 carbon. 3″, 4″ and 5″ carbon signals are observed at 108.2, 109.8 and 110.1 ppm respectively. In the HSQC spectrum, the H-4 proton signal correlates with the analogous carbon signal at 54.5 ppm. The H-3 and H-5 proton signal correlates with the analogous carbon signals at 57.0 and 52.4 ppm. The H-2 proton signal correlates with consequent carbon signal at 72.2 ppm. The 3″, 4″ and 5″ proton signals correlates with the corresponding carbon signals at 108.24, 109.85 and 110.15ppm. All the aromatic proton signals are correlated with the corresponding carbon signals. The $^1$H-$^{13}$C cosy correlations further confirmed the one dimensional $^1$H NMR spectral assignments.

Inhibitory Activity towards glucosamine-6-phosphate synthase (PDB ID: 2VF5)

Molecular docking studies were implementing to investigate the precise binding site of ligand on protein. The synthesized six analogues docked with antibacterial protein glucosamine-6-phosphate synthase. The Analogues 4a-g showed best ligand pose energy ranging from -11.06 to -13.45 kcal/mol in 2VF5. The 3D and 2D view of compound 4a with the standard drug (glucosamine-6-phosphosphate synthase) are shown in Fig. 1. The Best ligands pose energy, Van der Waals interaction, conventional hydrogen bond, pi-pi interaction, alkyl and pi-alkyl interaction of all the docked compounds including standard drug were presented in Table 1. The compound 4a showed very highest ligand pose energy -13.45 kcal/mol compared to glucosamine-6-phosphate (-6.31 kcal/mol). The analogue 4a is surrounded by Van der Waals, Alkyl, pi-alkyl and pi-loan pair interactions. ALA327, GLY301, LYS487, THR302 and GLU481 residues surrounded by Van der Waals interactions. The amino acid ASN305 interface in furfuryl ring exhibited Pi-Loan pair interaction with the bond distance of 4.42Å. CYS300, ILE326 and TYR304 amino acids accompanying benzoyl phenyl ring with alkyl and Pi- alkyl interactions. The amino acids LEU484 and LEU480 interact in furfuryl ring in alkyl and Pi-alkyl interactions. This ensured the binding affinity and results in an increased 2VF5 inhibitory activity.
But the standard drug molecules surrounded with Van der Waals, conventional hydrogen bond, carbon hydrogen bond with less amino acid interaction and also the bond distance is very high compared to analogue 4a. The compounds 4d, 4e and 4f containing halogen atoms also showed high ligand pose energy.

Figure 1(a). 3D and 2D view of binding interactions of compound

Figure 1(b). 3D and 2D view of binding interactions of standard drug 2VF5

Inhibitory activity towards human lanosterol 14-alpha dimethylase in complex with ketoconazole (PDB ID: 3LD6)

A docked 2D image of compound 4b with the standard molecule is shown in Fig. 2. The Best ligand pose energy, Van der Waals interaction, conventional hydrogen bond, pi-pi interaction, alkyl and pi-alkyl interaction of all the docked compounds including standard drug were presented in Table 2. The best ligand pose energy of compound 4b is -18.06 kcal/mol while the standard drug molecule is -10.47 kcal/mol. In the binding mode, the compound 4b was attractively bound to 3LD6 via Van der Waals interaction, pi-sigma, pi-lone pair, alkyl and pi-alkyl interactions. Compound 4b surrounded by Van der Waals interaction with the amino acid residues HIS489, ILE379, MET381, MET378, MET487, HIS236 and GLY78. One pi-sigma interaction with PHE105 residues in methyl substituted phenyl ring. The TRP758 amino acid interacts in alkyl interaction with indolone phenyl ring. TRP239 residues interface with one pi lone pair interaction in benzoyl carbonyl atom. Totally eight residues interact with alkyl and pi-alkyl interaction with the analogue 4b. Three residues like LYS103, VAL101 and LEU240 accompanying benzoylphenyl ring with
different bond distances. The remaining residues MET100, CYS402, PHE98 and TYR107 act together in methyl group and methyl substituted phenyl ring in different bond distances. The standard drug molecule is surrounded only less interactions like Van der Waals, carbon hydrogen bond, one pi-sigma, one pi-pi T shaped and alkyl and pi-alkyl interaction with fewer residues and also the bond distance is very high. This is confirmed the binding affinity and results in an increased 3LD6 inhibitory activity compared to standard drug molecule.

Figure 2(a). 3D and 2D view of binding interactions of compound

Figure 2(b). 3D and 2D view of binding interactions of standard drug 3LD6
Table 1. Different types of interactions in the compounds 4a-4g with 2VF5

<table>
<thead>
<tr>
<th>Compds.</th>
<th>Best ligand pose energy</th>
<th>Van der Waals interactions</th>
<th>Alkyl and Pi-alkyl interaction</th>
<th>Others</th>
</tr>
</thead>
<tbody>
<tr>
<td>4a</td>
<td>-13.45</td>
<td>ALA327, LY301, LYS487, HR302, GLU481</td>
<td>CYS300, ILE326, MET308, LEU480, LEU484</td>
<td>ASN305(Pi-loan pair)</td>
</tr>
<tr>
<td>4b</td>
<td>-11.77</td>
<td>GLU501, GLU495, LYS487</td>
<td>ALA498, TYR497, TYR476, ALA496, ALA483</td>
<td>TYR 497 (Pi-sigma), MET308 (Pi-sulfur)</td>
</tr>
<tr>
<td>4c</td>
<td>-11.06</td>
<td>ASN305, TYR304, ALA498, GLU495, ALA494</td>
<td>ALA483, LEU484, MET308, TYR476, LEU480, ALA496</td>
<td>LYS487, ALA496 (Conventional hydrogen bond) LEU484 (Pi-sigma) GLU495 (carbon hydrogen bond)</td>
</tr>
<tr>
<td>4d</td>
<td>-11.54</td>
<td>LEU484, THR302, GLU325, GLU329, MET308</td>
<td>-</td>
<td>LEU480 (Pi-sigma), GLY301(Halogen fluorine)</td>
</tr>
<tr>
<td>4e</td>
<td>-11.65</td>
<td>GLU487, GLU495, GLU501</td>
<td>ALA483, ALA496, LEU480, TYR497, ALA498, TYR476</td>
<td>MET308 (Pi-sulfur)</td>
</tr>
<tr>
<td>4f</td>
<td>-12.03</td>
<td>GLU495, TYR497, ALA498, GLY301, ASN305, LEU484</td>
<td>ALA496, TYR304, TYR476.</td>
<td>ILE326 (Pi-sigma) LYS487, (Conventional hydrogen bond)</td>
</tr>
<tr>
<td>4g</td>
<td>-10.95</td>
<td>ALA327, ALA299, MET308, GLU481</td>
<td>ALA483, ALA496, LEU484, LYS487, LEU480</td>
<td>GLY301, TYR476, ASN305 (Carbon and Pi-Donor hydrogen bond), ILE326 (Pi-sigma), CYS300 (Pi-sulfur)</td>
</tr>
<tr>
<td>Standard Drug</td>
<td>6.31</td>
<td>GLU524, TYR312</td>
<td>-</td>
<td>ASN522, ARG472 (Conventional hydrogen), ASP474, ARG472(salt bridge, attractive charge, pi-anion) GLY473 (carbon hydrogen bond)</td>
</tr>
</tbody>
</table>

Table 2. Different types of interaction in compounds 4a-g with 3LD6

<table>
<thead>
<tr>
<th>Compds.</th>
<th>Best ligand pose energy</th>
<th>Van der Waals interactions</th>
<th>Alkyl and Pi-alkyl interaction</th>
<th>Other interaction</th>
</tr>
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<td>4a</td>
<td>-16.62</td>
<td>LYS103, VAL101, PHE98, TRY131, LEU134</td>
<td>LEU240, MET487, MET381, TRP239, CYS402, ILE379, TYR107</td>
<td>MET100 (sulfur-(x)), PHE105 (Pi-sigma), PHE77, HIS236 (Pi-Donor hydrogen bond)</td>
</tr>
<tr>
<td>4b</td>
<td>-18.06</td>
<td>HIS489, ILE379, MET381, MET378, MET487, HIS236, GLY78</td>
<td>LYS103, VAL101, LEU240, TRP239, MET100, TYR107 CYS402, PHE98, HE77.</td>
<td>PHE105 (Pi-sigma), TRP239 (Pi-lone pair)</td>
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<tr>
<td>4c</td>
<td>-14.50</td>
<td>MET100, LEU240, LYS103, VAL101, VAL130, TYR131, PHE234, HIS489, MET378,</td>
<td>LEU134, ILE379, TYR107, LYS402, PHE77, PHE105, TRP239</td>
<td>-</td>
</tr>
<tr>
<td>4d</td>
<td>-13.81</td>
<td>MET381, ILE377, MET378, HIS489, ILE488, TYR145</td>
<td>ILE379, MET487, PHE234, LEU134</td>
<td>VAL130 (Conventional hydrogen bond), PHE234 (Pi-loan pair), TYR131</td>
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<tr>
<td>4e</td>
<td>-14.26</td>
<td>VAL130, MET378, HIS489, ILE377, ILE488, PHE139, TYR145</td>
<td>ILE379, MET487, PHE234, LEU134, MET381, TRP239, TYR131</td>
<td>PHE234 (Pi-sigma), TYR131 (Carbon hydrogen bond)</td>
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<tr>
<td>4f</td>
<td>-14.29</td>
<td>MET378, HIS489, HIS236, ILE488, PHE139, TYR145, MET381, ILE377</td>
<td>ILE379, MET487, PHE234, LEU134, TYR131</td>
<td>VAL130 (Conventional hydrogen bond), TYR131</td>
</tr>
<tr>
<td>4g</td>
<td>-13.57</td>
<td>MET378, HIS489, HIS236, ILE488, PHE139</td>
<td>LEU579, TRP684, TRY590, MET826, MET932, TYR59.</td>
<td>-</td>
</tr>
<tr>
<td>Standard Drug</td>
<td>-10.47</td>
<td>PRO526, PHE522, MET545, MET932, THR580, ILE933</td>
<td>MET826, TYR590, PHE679, TYR576</td>
<td>GLY523 (Carbon hydrogen bond), LEU579 (Pi-sigma)</td>
</tr>
</tbody>
</table>
Antimicrobial Studies

In vitro antibacterial activity

The synthesized compounds were screened for their antimicrobial activity against *K. pneumoniae*, *B. cereus*, *S. typhi*, *E. coli* and *V. cholerae* with the standard ciprofloxacin using an agar-well diffusion method for the determination of MIC (minimum inhibitory concentration of the compound). The compounds to be tested were dissolved in DMSO and different dilutions of the samples were prepared. A similar procedure was used for the commercial antibiotics streptomycin, which were used as positive controls for bacteria. The test organisms were transferred into freshly prepared Mueller-Hinton agar plates and evenly spread. After letting the plates dry for 5 minutes, wells were bored into the agar. 0.1 mL of each test compound and streptomycin (positive control) were added into the well, and then the plates were incubated at 37°C for 24 hours (for bacteria) and the zone of inhibition were measured. Compounds which showed a zone of inhibition less than 1.8 cm^{-1} were taken as a resistant, those with 1.8-2.0 cm^{-1} were taken as intermediate and those with zone of more than 2 cm^{-1} were assumed to be sensitive. Suitable dilutions of test compounds were added to 10 mL broth tubes inoculated with the test culture and incubated for 24 hours, such that their MIC activity was studied at 12, 15, 18 and 20 mg mL^{-1}. The tubes were checked for turbidity by comparing the positive control and negative control of the compounds against different strains of microorganism. It is confirmed from the in-vitro antimicrobial activity data of compounds 4a, 4b and 4g display the highest activity MIC 12.5 mg mL^{-1} against *Klebsilae pneumonia* while 4c, 4d, 4e and 4f respectively, showed somewhat inferior activity compared to the standard streptomycin (MIC 12.5 mg mL^{-1}). In the case of *B. Cereus*, compounds 4a, 4b, 4c and 4f were found to be the most active derivative in vitro with a MIC of 12.5, 25, 25 and 12.5 mg mL^{-1}, respectively. Compounds 4a and 4b showed strong activity with MIC 12.5, mg mL^{-1} against *S.typhi*. However, the compounds 4c, 4d, 4e and 4f showed good to moderate activity whereas compound 4g did not show the activity in the concentration range tested. In the case of *E. coli*, compound 4e showed good to moderate activity with MIC 25 mg mL^{-1}. Further compounds 4a, 4e and 4f showed good activity against *V. cholerae* whereas the compounds 4b, 4c, 4d and 4e showed moderate activity as compared to the standard antimicrobial agent streptomycin (MIC 12.5 mg mL^{-1}). The compounds 4e and 4f showed good activity against almost the majority of the bacterial strains, whereas 4b, 4c and 4g showed weak activity against the majority of the bacterial strains. The tendency of the antibacterial activity of the synthesized compounds is as follows: 4a > 4e > 4f > 4b > 4c > 4d > 4g. Minimum inhibitory concentration of antibacterial activities is shown in Table 3. The Bar diagram of MIC values of antibacterial activities are shown in Fig. 3.

In vitro antifungal activity

3-benzoyl-5-(furan-2-yl)-4-phenylspiro[indoline-3,2-pyrrolidin]-2-one derivatives 4a-g were also screened for their in-vitro antifungal activity with fungal strains viz., *C. albicans*, *A. niger*, *A. flavus*, *C. neoformans* and *F. oxysporum*. Here, Ketoconazole be used as the standard drug. MIC character ranging from 6.25-12.5 mg/mL is listed in Table 4. Compound 4b (MIC= 3.125 lg/mL) methyl substituent in the para position of phenyl ring in C-4 carbon at five membered pyrrole heterocyclic ring showed superior antifungal activity against *C. albicans* and *A. niger*. Compound 4b (MIC= 3.125 mg/mL) left out any substituent in the para position of phenyl ring in C-4 carbon at pyrrole five membered heterocyclic ring exhibited excellent antifungal action than the standard drug against *C. albicans*. Compound 4c, showed equivalent activity against *C. albicans*, *A. niger* and *A. flavus* fungal strains. The compounds 4d, 4e and 4f showed excellent activity against almost all the fungal strains except *C. neoformans* and *F. oxysporum*. Originally, the tendency of the antifungal activity of all the compounds is as follows: 4b> 4a > 4c > 4d > 4e > 4f > 4g. The Bar diagram of MIC values of antifungal activities are shown in Fig. 4.
**Figure 3.** Minimum inhibitory concentrations of antibacterial activity of compounds 4a-4g.

**Figure 4.** Minimum inhibitory concentrations of antifungal activity of compounds 4a-4g

**Table 3.** Minimum inhibitory concentration (µg ml⁻¹) of antibacterial activity of compounds 4a-g

<table>
<thead>
<tr>
<th>Compds.</th>
<th><strong>K. pneumonia</strong></th>
<th><strong>B. cerus</strong></th>
<th><strong>S. typhi</strong></th>
<th><strong>E. coli</strong></th>
<th><strong>V. cholerae</strong></th>
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<tbody>
<tr>
<td>4a</td>
<td>12.5</td>
<td>12.5</td>
<td>12.5</td>
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<td>4c</td>
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<td>12.5</td>
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Table 4. Minimum inhibitory concentration (µg ml⁻¹) of antifungal activity of compounds 4a-g

<table>
<thead>
<tr>
<th>Compds.</th>
<th>Minimum inhibitory concentration (µg ml⁻¹)</th>
</tr>
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<tr>
<td></td>
<td>C. albicans</td>
</tr>
<tr>
<td>4a</td>
<td>3.125</td>
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<tr>
<td>4b</td>
<td>3.125</td>
</tr>
<tr>
<td>4c</td>
<td>6.25</td>
</tr>
<tr>
<td>4d</td>
<td>6.25</td>
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<tr>
<td>4e</td>
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<td>4f</td>
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<td>4g</td>
<td>25</td>
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<td>Ketaconazole</td>
<td>6.25</td>
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3'-benzoyl-5'-((furan-2-yl)-4'-(p-tolyl)spiro[indoline-3,2'-pyrrolidin]-2-one (4a)
M.F.: C₂₈H₂₂N₂O₃, m.p. (°C): 248-251, Yield(%): 84, IR (Neat, cm⁻¹); 1714.80, 1700.62 (C=O), 3250.50, 3110.84 (NH). ¹H NMR (CDCl₃, ppm); δ: 4.55 (d, H-5, J= 8.4Hz), 5.54 (s, H-3, H-4), 6.00 (s, 1H, 3'' furan), 6.12 (s, 1H, 4'' furan), 6.67 (d, 1H, J=6.4Hz, 5'' furan), 6.91-7.91 (m, 14H Aromatic protons). ¹³C NMR (CDCl₃, ppm); δ: 196.5 (C=O), 180.3 (C=O), 153.7 (C-O, furan), 72.2 (C-2), 57.0 (C-3), 54.5 (C-5), 52.4 (C-4), 108.2 (3'' carbon), 110.9 (4'' carbon), 110.3 (5'' carbon), 123.3, 123.9, 127.7, 128.3, 128.4, 128.5, 128.8, 129.2, 129.4, 129.5, 130.0, 133.2, 133.4, 137.1, 140.7, 142.4, 142.5 (Aromatic carbon).

3'-benzoyl-5'-((furan-2-yl)-4'-(4-methoxyphenyl)spiro[indoline-3,2'-pyrrolidin]-2-one (4b)
M.F.: C₂₉H₂₄N₂O₄, m.p. (°C): 236-242, Yield(%): 76, IR (Neat, cm⁻¹); 1710.23, 1708.42 (C=O), 3248.00, 3112.04 (NH). ¹H NMR (CDCl₃, ppm); δ: 3.87 (s, OCH₃), 4.62 (d, H-5, J= 13.2 Hz), 5.56 (s, H-3, H-4), 6.03 (d, J=3.2 Hz, 1H, 3'' furan), 6.13 (d, J=5.2Hz, 1H, 4'' furan), 6.86 (d, 1H, J=6.4Hz, 5'' furan), 6.87-7.92 (m, 14H Aromatic protons). ¹³C NMR (CDCl₃, ppm); δ: 196.8 (C=O), 180.9 (C=O), 158.7 (C-O, furan), 72.4 (C-2), 61.1 (C-3), 57.0 (C-5), 52.3 (C-4), 21.0 (CH₃), 108.1 (3'' carbon), 109.6 (4'' carbon), 110.1 (5'' carbon), 123.3, 123.9, 127.6, 127.7, 127.8, 127.9, 128.1, 128.2, 128.3, 128.4, 128.5, 128.8, 129.4, 129.5, 130.3, 133.0, 137.3, 140.1, 140.9, 142.4 (Aromatic carbon).

3'-benzoyl-5'-((furan-2-yl)-4'-(4-fluorophenyl)spiro[indoline-3,2'-pyrrolidin]-2-one (4c)
M.F.: C₂ₘH₂₁FN₂O₃, m.p. (°C): 236-239, Yield(%): 78, IR (Neat, cm⁻¹); 1709.67, 1710.19 (C=O), 3248.05, 3116.03 (NH). ¹H NMR (CDCl₃, ppm); δ: 4.68 (d, H-5, J= 8.4Hz), 5.57 (s, H-3, H-4), 6.03 (d, J=2.8Hz, 1H, 3'' furan), 6.13 (d, J=5.2Hz, 1H, 4'' furan), 6.86 (d, 1H, J= 8.8 Hz, 5'' furan), 6.87-7.92 (m, 14H Aromatic protons). ¹³C NMR (CDCl₃, ppm); δ: 196.6 (C=O), 180.9 (C=O), 154.8 (C-O, furan), 72.4 (C-2), 62.3 (C-3), 57.0 (C-5), 52.4 (C-4), 55.0 (OCH₃), 107.9 (3'' carbon), 109.7 (4'' carbon), 110.1(5'' carbon), 113.3, 113.5, 114.1, 114.5, 119.8, 122.5, 123.1, 123.9, 124.5, 125.8, 126.7, 128.3, 128.5, 128.8, 130.3, 133.0, 137.3, 140.1, 140.9, 142.3 (Aromatic carbon).

3'-benzoyl-4'-(4-fluorophenyl)-5'-(furan-2-yl)spiro[indoline-3,2'-pyrrolidin]-2-one (4d)
M.F.: C₂₉H₂₁FN₂O₃, m.p. (°C): 226-230, Yield(%): 80, IR (Neat, cm⁻¹); 1713.00, 1718.61 (C=O), 3242.51, 3110.02 (NH). ¹H NMR (CDCl₃, ppm); δ: 4.68 (d, H-5, J= 8.4Hz), 5.57 (s, H-3, H-4), 6.03 (s, 1H, 3'' furan), 6.18 (s, 1H, 4'' furan), 6.73 (d, 1H, J=6.4Hz, 5'' furan), 6.96-8.03 (m, 14H Aromatic protons). ¹³C NMR (CDCl₃, ppm); δ: 196.6 (C=O), 180.5 (C=O), 153.8 (C-O, furan), 72.3 (C-2), 57.0 (C-3), 54.4 (C-5), 52.4 (C-4), 108.2 (3'' carbon), 109.7 (4'' carbon), 110.2 (5'' carbon), 114.9, 115.1, 116.1, 116.3, 121.8, 123.2, 123.9, 127.7, 128.1, 128.3, 128.5, 128.6, 129.1, 129.3, 129.4, 129.6, 130.1, 130.3, 130.4, 132.9, 133.0, 133.2, 137.2, 138.1, 140.7, 142.4, 143.6 (Aromatic carbon).
3'-benzoyl-4'-(4-chlorophenyl)-5'-furan-2-yl)spiro[indoline-3,2'-pyrrolidin]-2-one (4e)
M.F.: C_{28}H_{21}ClN_{2}O_{3}, m.p.(0°C): 240-243, Yield (%): 81, IR (Neat, cm^{-1}): 1710.00, 1701.62 (C=O), 3260.52, 3109.01 (NH). 1H NMR (CDCl₃, ppm); δ: 4.68 (d, H-5, J= 12.4 Hz), 5.57 (s, H-3, H-4), 6.03 (d, J=3.2 Hz, 1H, 3″ furan), 6.19 (s, 1H, 4″ furan), 6.96 (d, 1H, J= 8.8 Hz, 5″ furan), 7.25 - 8.03 (m, 14H Aromatic protons). 13C NMR (CDCl₃, ppm); δ: 190.7 (C=O), 179.5 (C-O, furan), 72.4 (C-2), 58.5 (C-3), 57.1 (C-5), 55.2 (C-4), 108.8 (3″ carbon), 109.7 (4″ carbon), 110.3 (5″ carbon), 122.1, 127.8, 127.9, 128.0, 128.1, 128.3, 128.6, 130.6, 132.8, 134.9, 138.2, 142.3, 144.9 (Aromatic carbon).

3'-benzoyl-4'- (4-bromophenyl)-5'-furan-2-yl)spiro[indoline-3,2'-pyrrolidin]-2-one (4f)
M.F.: C_{28}H_{21}BrN_{2}O_{3}, m.p. (0°C) : 254-257, Yield(%): 73, IR (Neat, cm^{-1}); 1710.60, 1701.92 (C=O), 3248.03, 3104.00 (NH). 1H NMR (CDCl₃, ppm); δ: 4.62 (d, H-5, J= 12.4 Hz), 5.56 (s, H-3, H-4), 6.03 (d, J=3.2 Hz, 1H, 3″ furan), 6.13 (s, 1H, 4″ furan), 6.86 (d, 1H, J= 8.8 Hz, 5″ furan), 6.87 -7.92 (m, 14H Aromatic protons). 13C NMR (CDCl₃, ppm); δ: 200.1 (C=O), 196.5 (C=O), 153.7 (C-O, furan), 72.1 (C-2), 57.0 (C-3), 54.6 (C-5), 52.4 (C-4), 108.2 (3″ carbon), 109.8 (4″ carbon), 110.3 (5″ carbon), 121.5, 123.3, 123.9, 127.7, 128.1, 128.3, 128.5, 128.6, 128.7, 128.7, 129.5, 129.6, 129.8, 131.0, 131.3, 131.8, 133.2, 133.9, 140.7, 142.4 (Aromatic carbon).

3'-benzoyl-4'-(4-(dimethylamino)phenyl)-5'-furan-2-yl)spiro[indoline-3,2'-pyrrolidin]-2-one (4g)
M.F.: C_{30}H_{27}N_{3}O_{3}, m.p. (0°C) : 251-253, Yield(%): 71, IR (Neat, cm^{-1}); 1711.70, 1701.22 (C=O), 3248.03, 3104.00 (NH). 1H NMR (CDCl₃, ppm); δ: 3.01 (s, 6- H, (NCH₃)₂, 4.68 (d, H-5, J= 12.4 Hz), 5.57 (s, H-3, H-4), 6.03 (d, J=3.2 Hz, 1H, 3″ furan), 6.18 (s, 1H, 4″ furan), 6.73 (d, 1H, J= 8.8 Hz, 5″ furan), 6.96-8.03 (m, 14H Aromatic protons). 13C NMR (CDCl₃, ppm); δ: 196.4 (C=O), 180.9 (C=O), 158.7 (C-O, furan), 72.4 (C-2), 57.8 (C-3), 54.4 (C-5), 52.4 (C-4), 62.3 (N-CH₃)₂, 107.9 (3″ carbon), 109.8 (4″ carbon), 110.3 (5″ carbon), 121.5, 123.3, 123.9, 127.7, 128.1, 128.3, 128.5, 128.6, 128.7, 128.7, 129.5, 129.6, 129.8, 131.0, 131.3, 131.8, 133.2, 133.9, 140.7, 142.4 (Aromatic and ipso carbon).

Conclusions

3'-benzoyl-5'-furan-2-yl)-4'-phenylspiro[indoline-3,2'-pyrrolidin]-2-one derivatives (4a-g) have been synthesized and characterized by IR, 1H and 13C NMR spectral techniques. In addition, two-dimensional NMR spectra of 1H-13C COSY spectrum have been recorded for 4a. Molecular docking studies were carried out to investigate the precise binding site of ligand on protein. The compound 4a showed very high ligand pose energy (-13.45 kcal/mol) compared to glucosamine-6-phosphate (-6.31 kcal/mol). The best ligand pose energy of compound 4b is -18.06 kcal/mol whereas the standard drug molecule is -10.47 kcal/mol. In this binding mode, compound 4b was attractively bound to 3LD6 via Vander Waals interactions. The in vitro antimicrobial activity data of compounds 4a, 4b and 4g displayed the highest activity (MIC 12.5 µg/ml - against Klebsilae pneumonia and in the case of B. Cereus, compounds 4a, 4b, 4c and 4f were found to be the most active derivative in vitro with a MIC of 12.5, 25, 25 and 12.5 µg/ml, respectively. Compound 4b (MIC= 3.125 µg/ml) exhibited superior antifungal activity against C. albicans and A. niger related to other fungal strains with compared to standard amphotericin B.

Conflict of Interest

The authors declare that there is no conflict of interest.

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References


